# **Supplementary Information**

## Aluminium fluorescence detection with a FRET amplified chemosensor

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#### **1. Experimental Procedures.**

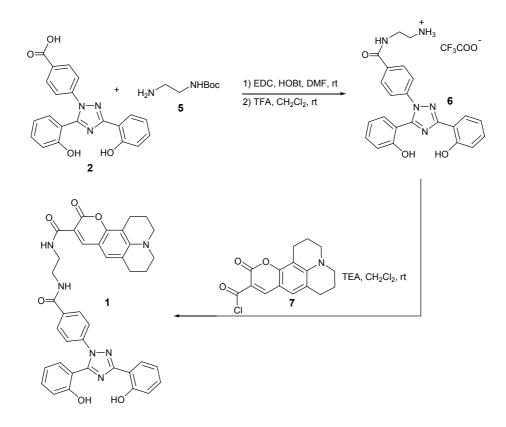
**General:** <sup>1</sup>H NMR spectra were run on a Jeol EX-400 instrument. Chemical shifts are reported relative to internal Me<sub>4</sub>Si. Multiplicity is given as usual. ESI-MS spectra were obtained on a PE-API spectrometer at 5600 volts by infusion of methanolic solutions. UV-Vis absorption measurements were performed on a Perkin Elmer Lambda 16 spectrophotometer equipped with a thermostated cell holder. Fluorescence spectra were recorded on a Perkin Elmer LS-50B spectrometer equipped with a thermostated cell holder. TLC's were performed on Polygram<sup>®</sup> Sil G/UV<sub>254</sub> silica gel pre-coated plastic sheets. Flash chromatography was run on silica gel, 230-400 mesh ASTM (Kieselgel 60, Merck). Solvents were purified under standard techniques. Reagents were purchased by Aldrich and used as received. Al(NO<sub>3</sub>)<sub>3</sub>, Cu(NO<sub>3</sub>)<sub>2</sub>, Zn(NO<sub>3</sub>)<sub>2</sub>, Ni(NO<sub>3</sub>)<sub>2</sub>, FeCl<sub>3</sub>, CaCl<sub>2</sub>, MgCl<sub>2</sub> were analytical grade products. Metal ion stock solutions were titrated against EDTA following standard procedures. 4-[3,5-bis-(2-hydroxyphenyl)-1,2,4-triazol-1-yl]-benzoic acid<sup>1</sup> (**2**), N-BOC-1,2-diaminoethane<sup>2</sup> (**5**) were prepared by known procedures.

**Spectrometric titrations:** Several 2-mL solutions of buffer  $(1 \cdot 10^{-2} \text{ M})$ , ligands **1** or **3**  $(3.11 \cdot 10^{-6} \text{ M})$  in the case of fluorescence experiments or  $0.9 \cdot 1.6 \cdot 10^{-5} \text{ M}$  in the case of the absorption experiments) and the desired amount of metal salt solution in water/ethanol 1:1 were prepared in plastic fluorescence cuvettes and incubated at 25° C for 12 hours. Complex formation was monitored following the absorption spectral changes. UV-Vis or fluorescence spectra were then recorded. From the spectral changes observed upon addition of the substrate, the K<sub>app</sub> values were obtained by non-linear regression analyses of fluorescence data (at the selected wavelength) versus metal ion concentrations. The buffer used in the experiments was acetate buffer 0.01 M at pH 4.0. After mixing with ethanol the pH value read with the pH-meter was 5.0.

<sup>&</sup>lt;sup>1</sup>R.Lattmann, P.Acklin (Novartis AG), WO-A 9749395A1 1997 [Chem.Abstr. 1998, **128**, 114953e] <sup>2</sup>P.Krapcho, C.S.Kuell, *Synth.Commun*. 1990, **20**, 2559-2564.

#### 2. Synthesis

#### 2.1 Synthesis of derivative 1



A CH<sub>2</sub>Cl<sub>2</sub> solution of HOBt (0.36g, 2.7 mmol) was added at r.t. to a solution of **2** (4-[3,5-bis-(2-hydroxyphenyl)-1,2,4-triazol-1-yl]-benzoic acid) (1g, 2.7 mmol) in 10 ml anhydrous DMF. After cooling down the solution to 0°C, EDC (0.062g, 3.2 mmol) was added, the mixture stirred for further 10 min at r.t., then a solution of **5** (mono <sup>t</sup>butyloxycarbonyl-1,2-ethylenediamine) (0.43g, 2.7 mmol) and Et<sub>3</sub>N (0.4ml, 3.2 mmol) in 10ml CH<sub>2</sub>Cl<sub>2</sub> was added dropwise. The mixture was left on stirring, monitoring the course of the reaction by TLC (CHCl<sub>3</sub>:EtOH 15:1). After 18h, the solution was diluted with CH<sub>2</sub>Cl<sub>2</sub> (20ml), washed with 5% citric acid (3x) and water (5x). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>), the solvent removed *in vacuo*, and the crude material was purified by FC (eluent: toluene-ethyl acetate 7:3) to give the product **6**, 0.260g, yield 19%.

<sup>1</sup>H-NMR (400 MHz, MeOD) :  $\delta$  1.49 (s, 9H), 3.34 (t, 2H), 3.52 (t, 2H), 6.91 (d, 1H, J=8.0 Hz), 7.06 (m, 3H), 7.44 (m, 2H), 7.56 (dd, 1H, J = 7.7, 1.5 Hz), 7.62, 7.95 (AB quartet, 4H), 8.19 (dd, 1H, J 7.7, 1.5 Hz). ESI-MS calcd for C<sub>28</sub>H<sub>29</sub>N<sub>5</sub>O<sub>5</sub> (M<sup>+</sup>): 515; obsd: 538 (M<sup>+</sup> + Na<sup>+</sup>).

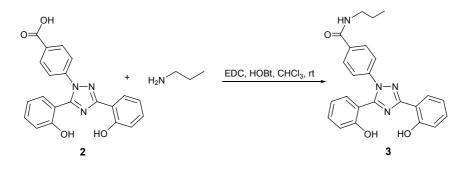
A solution of above Boc-protected derivative, (0.260g, 0.5 mmol) and TFA (10 ml) in CH<sub>2</sub>Cl<sub>2</sub> (10ml) was stirred 1h at r.t. Evaporation of the solvent to dryness left a residue (product **6**) which was used without further purification. <sup>1</sup>H-NMR (400 MHz, MeOD),  $\delta$  : 3.25 (t, 2H), 3.75 (t, 2H), 6.93 (d, 1H J = 8.4 Hz), 7.07 (m, 3H), 7.44 (m, 2H), 7.54 (dd, 1H J=7.3,1.5 Hz), 7.65, 8.00 (AB

quartet, 4H), 8.18 (dd, 1H , J=7.7, 1.5 Hz). ESI-MS calcd for  $C_{23}H_{21}N_5O_3$  (M<sup>+</sup>): 415; obsd: 438 (M<sup>+</sup> + Na<sup>+</sup>).

Oxalylchloride (0.1 ml) and DMF (10  $\mu$ l) were added under Ar to a CH<sub>2</sub>Cl<sub>2</sub> solution (10 ml) of Coumarin 343 (0.050 g 0.17 mmol). The solution was stirred at r.t. 1h, then the solvent was removed *in vacuo*. The resulting acid chloride 7<sup>3</sup> (0.050g, 0.16 mmol), dissolved in 10 ml CH<sub>2</sub>Cl<sub>2</sub>, was dropwise added to a solution of **6** (0.086 g , 0.16 mmol) and Et<sub>3</sub>N (0.075 ml, 0.52 mmol) in the same solvent. After sirring at room temperature 18h, the solution was extracted with H<sub>2</sub>O, the organic phase dried and the solvent evaporated *in vacuo*. Purification by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 10:1) gave the product **1**, 0.075 g, 67% yield.

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.97 (m, 4H), 2.76 (t, 2H), 2.89 (t, 2H), 3.34 (m, 4H), 3.71 (bt, 2H), 3.76 (bt, 2H), 6.65 (t, 1H), 6.94 (dd, 1H J = 8.0, 1.5 Hz), 7.00 (s, 1H), 7.06 (d, J = 8.0 Hz, 1H), 7.08 (d, J = 8.04 Hz, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.33 (m, 1H), 7.38 (m, 1H), 7.60, 8.10 (AB quartet, 4H), 8.14 (dd, 1H, J=8.0, 1.8 Hz), 8.38 (bt, 1H), 8.61 (s, 1H), 9.38 (t, 1H), 9.64 (s, 1H), 11.44 (s, 1H). ESI-MS calcd for C<sub>39</sub>H<sub>34</sub>N<sub>6</sub>O<sub>6</sub> (M<sup>+</sup>): 682; obsd: 705 (M<sup>+</sup> + Na<sup>+</sup>).

#### 2.2 Synthesis of ligand derivative 3

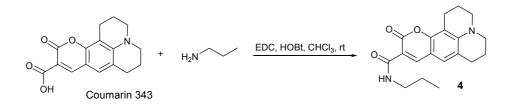


**2** (60 mg, 0.16 mmol) was suspended in 4 ml of CHCl<sub>3</sub>, triethylamine (0.10 mL, 0.72 mmol) and hydroxybenzotriazole (58 mg, 0.43 mmol) were added and the reaction mixture was cooled to 0° C. Then EDC·HCl (80 mg, 0.42 mmol) was added and, after 10 minutes, *n*-propylamine (0.020 mL, 0.47 mmol). The reaction mixture was stirred at 0° C for 1 hour and at room temperature overnight. After this time, 100 mL of CH<sub>2</sub>Cl<sub>2</sub> were added and the mixture was extracted with a 5% KHSO<sub>4</sub> solution (2x), a 5% NaHCO<sub>3</sub> solution (2x) and water (2x). The organic phase was dried (NaSO<sub>4</sub>) and the solvent removed to yield 62 mg (94%) of **3** as a white solid.

<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ : 1.03 (t, J = 7.2 Hz, 3H), 1.68 (m, 2 H), 3.47 (t, J = 6.3 Hz, 2H), 6.2 (bs, 1H), 6.6 (t, J = 7.0 Hz, 1H), 7.4-6.9 (m, 6H), 7.60 (d, J = 8.1 Hz, 2H), 7.96 (d, J = 8.1 Hz, 2H), 8.16 (d, J = 7.0 Hz, 1H). ESI-MS calcd for C<sub>24</sub>H<sub>22</sub>N<sub>4</sub>O<sub>3</sub> (M<sup>+</sup>): 414; obsd: 437 (M<sup>+</sup> + Na<sup>+</sup>).

<sup>&</sup>lt;sup>3</sup> J Gompel, G.Schuster, J.Org.Chem. 1987, **52**, 1465-1468.

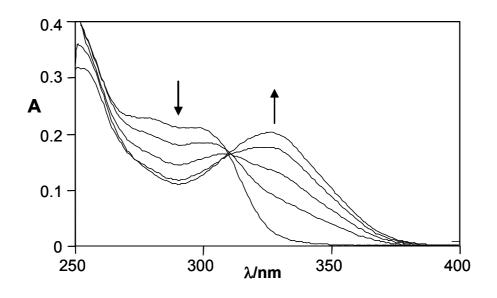
#### 2.2 Synthesis of coumarine derivative 4



Coumarin 343 (30 mg, 0.11 mmol), triethylamine (0.70 mL, 0.49 mmol) and hydroxybenzotriazole (43 mg, 0.32 mmol) were dissolved in 2 ml of CHCl<sub>3</sub> and the reaction mixture was cooled to 0° C. Then EDC·HCl (64 mg, 0.32 mmol) was added and, after 10 minutes, *n*-propylamine (0.014 mL, 0.32 mmol). The reaction mixture was stirred at 0° C for 1 hour and at room temperature overnight. After this time, 100 mL of CH<sub>2</sub>Cl<sub>2</sub> were added and the mixture was extracted with a 5% KHSO<sub>4</sub> solution (2x), a 5% NaHCO<sub>3</sub> solution (2x) and water (2x). The organic phase was dried (Na<sub>2</sub>SO<sub>4</sub>) and the solvent removed. The crude product was purified by flash column chromatography (silica gel, CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH 20:1) to yield 35 mg (97%) of **4** as a white solid.

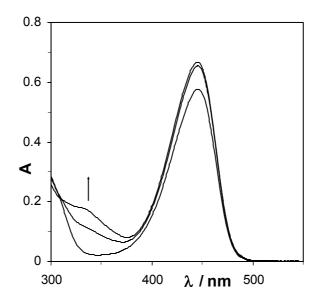
<sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ : 0.97 (t, J = 7.5 Hz, 3H), 1.62 (m, 2 H), 1.96 (m, 4H), 2.76 (t, J = 6.3 Hz, 2H), 2.87 (t, J = 6.5 Hz, 2H), 3.3-3.4 (m, 6H), 7.03 (s, 1H), 8.64 (s, 1H), 9.04 (bs, 1H). ESI-MS calcd for C<sub>19</sub>H<sub>22</sub>N<sub>2</sub>O<sub>3</sub> (M<sup>+</sup>): 326; obsd: 349 (M<sup>+</sup> + Na<sup>+</sup>).

## **3.** UV-Visible titration of ligand **3** with Al<sup>3+</sup>



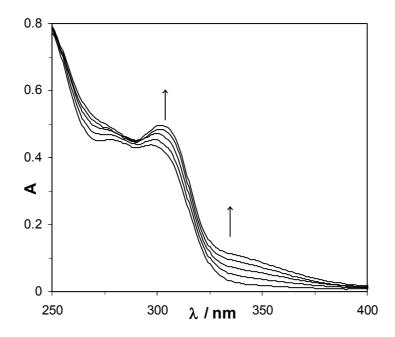
**Figure S1:** UV-Visible spectra of ligand **3** in the presence of increasing amounts of Al(NO<sub>3</sub>)<sub>3</sub> in EtOH/H<sub>2</sub>O (1/1) at pH= 5.0. [**3**]=  $8.7 \times 10^{-6}$  M, [acetate buffer] = 0.01M.

## 4. UV-Visible titration of compound 1 with Al<sup>3+</sup>

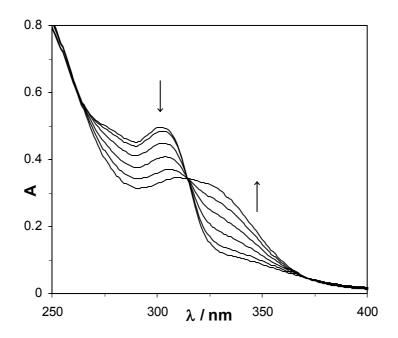


**Figure S2:** UV-Visible spectra of compound 1 in the presence of increasing amounts of Al(NO<sub>3</sub>)<sub>3</sub> in EtOH/H<sub>2</sub>O (1/1) at pH= 5.0. [1]=  $1.65 \times 10^{-5}$  M, [acetate buffer] = 0.01M.

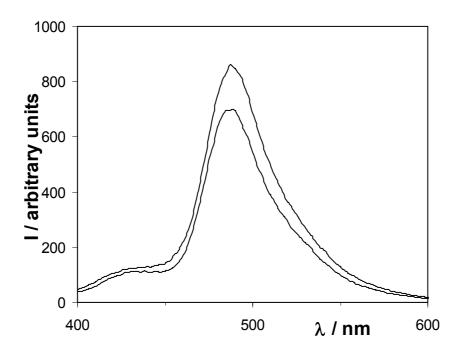
### 5. UV-Visible spectra of ligand 3 at increasing pH values



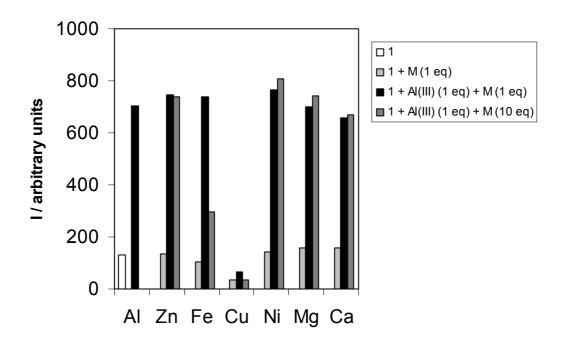
**Figure S3:** UV-Visible spectra of ligand **3** ( $1.74 \cdot 10^{-5}$  M) at increasing pH values in the range 7-12 in EtOH/H<sub>2</sub>O 1:1.



**Figure S4:** UV-Visible spectra of ligand **3** ( $1.74 \cdot 10^{-5}$  M) at increasing pH values in the range 12-13.9 in EtOH/H<sub>2</sub>O 1:1.



**Figure S5:** Fluorescence emission spectra ( $\lambda_{exc}$ = 445 nm) of sensor 1 (upper curve) and coumarin derivative 4 (lower curve) in ethanol. [1] = [4] = 3.11 \cdot 10^{-6} M.



**Figure S6:** Fluorescence response of chemosensor **1** to  $Al^{3+}$  in the presence of other metal ions. Conditions: EtOH/H<sub>2</sub>O (1/1), pH= 5.0. [**1**]=  $3.0x10^{-6}$  M, [acetate buffer] = 0.01M.